Corneal Scraping for Acanthamoeba

Arrangements

Follow the special instruction listed below, if you have any questions please contact the F.C. Blodi Eye Pathology Laboratory (319-335-7095) prior to beginning this procedure.

Make sure the following materials are in the room prior to performing the scraping:

i. If you are using the Saccomanno or cytology Fixative
   a) Spatula for scraping the cornea
   b) Sterile container (centrifuge tube, specimen container)
   c) Wash bottle containing Saccomanno fixative
   d) Ocular Pathology Consultation Request form with the proper basic patient information, clinical data, and specimen location (i.e: corneal scrapping right eye) written on the form.

ii. If you are using the 10% Neutral buffered formalin method:
   a) Spatula for scraping the cornea
   b) Slides with one end frosted
   c) Coplin jar with 10% neutral buffered formalin
   d) Paper toweling

The following procedural instructions based on the materials available to your institution are:

i. If you have Saccomanno or cytology fixative available use these instructions (this is the preferred method):
   a) Have an open sterile container (centrifuge tube, specimen container) available prior to scraping the patient.
   b) Using a spatula scrape the patient as usual.
   c) Immediately after scraping the patient, using wash bottle containing fixative, wash cells and tissue off spatula and into the sterile container.
   d) Make sure not to use more than 0.5 ml of fixative to wash the cells off the spatula.
   e) Past experience has shown when more than 0.5 ml of fixative is used, the lab produces 12 cytospin slides. There is usually a sparse cell count per slide. When using 0.5 ml Saccomanno, the lab produces 6 slides with a better cell representation per slide.
   f) Take tip of spatula and swirl in the Saccomanno to assure that all of the cells have been removed from the spatula.
   g) Place the lid on the container tightly
   h) Label the specimen container with the patient’s name and date of birth

ii. If you are using 10% Neutral Buffered Formalin:
   a) Scrape the cornea, immediately smear the scrapings on the clear portion of a clean glass slide. The specimen should be smeared so that the frosted end is facing up.
   b) Immediately immerse the slide containing the specimen in the coplin jar containing 10% Neutral Buffered Formalin.
   c) Allow the slides to sit in the 10% Neutral Buffered Formalin fixative for 15-20 minutes.
Note: **DO NOT** leave the slides in the formalin more than 15-20 minutes or the tissue may fall off the slides.

- Remove the slides from the fixative.
- Place the slides on a paper towel to air dry.

Note: Make sure the side of the slide containing the specimen is right side up or the scraping may be wiped off.

- After **completely** air dried, place the slides into a slide mailer (can be provided by the F.C. Blodi Eye Pathology laboratory).
- Fill out the Ocular Pathology Consultation Request form **completely**.